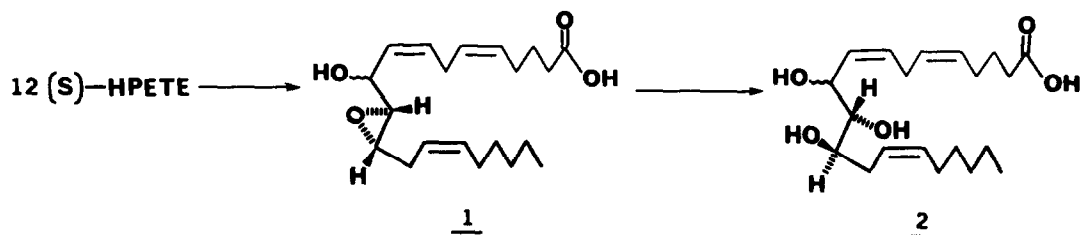


### SYNTHESIS OF TRIOXILIN B<sub>3</sub>

Sun Lumln, Pendri Yadagiri, and J. R. Falck\*  
Departments of Molecular Genetics and Pharmacology  
University of Texas Southwestern Medical Center  
Dallas, Texas 75235 USA

**Summary:** Both C(10) diastereomers of trioxilin B<sub>3</sub>, presumed to be a mixture of 10(R/S), 11(R), 12(R)-trihydroxyeicosa-5(Z), 8(Z), 14(Z)-trienoic acids, were prepared from a carbohydrate derived precursor by Wittig homologation and Mitsunobu inversion.

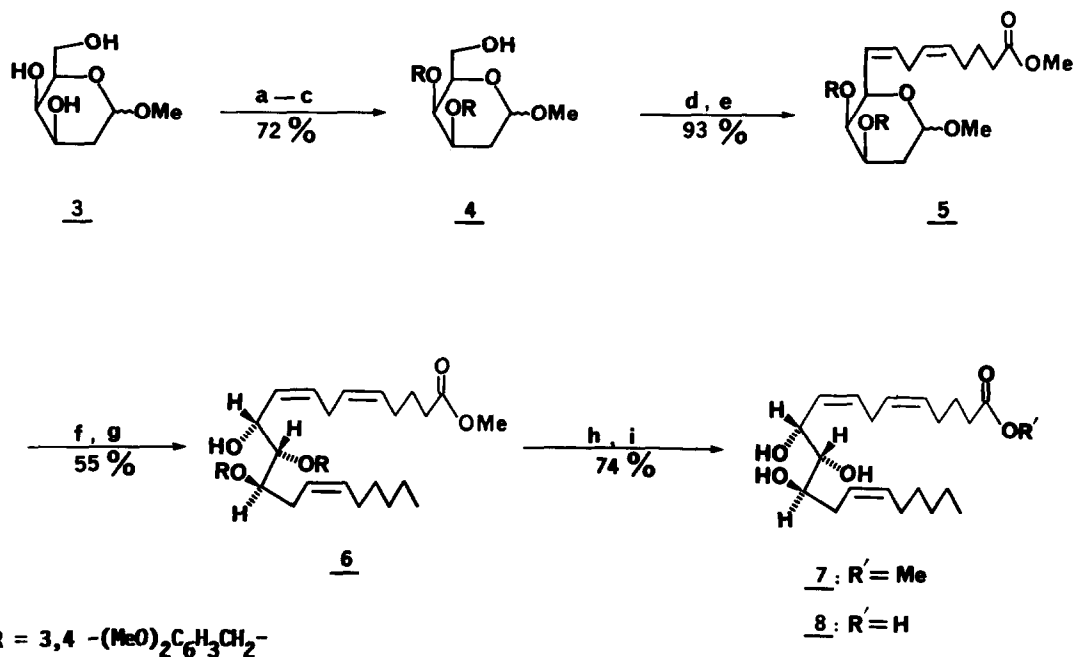
Hepoxilin B<sub>3</sub>, first identified by Walker et al<sup>1</sup> in 1979, arises via intramolecular rearrangement<sup>2</sup> of 12(S)-hydroperoxyeicosatetraenoic acid [12(S)-HPETE] and is now known to consist of two isomeric 10-hydroxy-11,12-epoxyeicosatrienoic acids (**1**) whose exact stereochemical constitution has been confirmed by total chemical synthesis<sup>3</sup>. Stable isotope studies<sup>4</sup> revealed **1** is regiospecifically hydrated at C(12) to the corresponding triols **2**, designated trioxilin B<sub>3</sub>, by an epoxide hydratase present in rat lung homogenate. Based on the well precedented<sup>5</sup> expectation of stereochemical inversion at the site of hydration, **2** can be formulated as 10(R/S), 11(R), 12(R)-trihydroxyeicosa-5(Z), 8(Z), 14(Z)-trienoic acid.



Interest in the arachidonate metabolites of the hepoxilin/trioxilin pathway has been heightened recently by reports of their insulin secretagogue activity<sup>6</sup> and possible role as second messengers for presynaptic inhibition of *Aplysia* sensory cells<sup>7</sup>. Related metabolites originating from linoleic, linolenic, eicosatrienoic, and eicosapentaenoic

acids have been observed previously, although generally their stereochemistries remain obscure<sup>8</sup>. To expedite the further study of the occurrence and pharmacological profile of this novel class of eicosanoids, we report herein the enantiospecific total synthesis of both C(10) diastereomers of trioxilin B<sub>3</sub>.

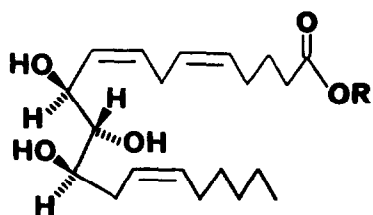
Scheme I



<sup>a</sup>Ph<sub>3</sub>CCl, CH<sub>2</sub>Cl<sub>2</sub>/C<sub>5</sub>H<sub>5</sub>N (8:1), DMAP, 40°C, 12h. <sup>b</sup>3,4-(MeO)<sub>2</sub>C<sub>6</sub>H<sub>3</sub>CH<sub>2</sub>Cl, KH, THF, 45°C, 3h. <sup>c</sup>ZnBr<sub>2</sub> (5 equiv), CH<sub>2</sub>Cl<sub>2</sub>/MeOH (20:1), 24°C, 2h. <sup>d</sup>(COCl)<sub>2</sub>, DMSO, CH<sub>2</sub>Cl<sub>2</sub>, -78°C, 1h; Et<sub>3</sub>N, -20°C, 0.5h. <sup>e</sup>9, THF/HMPA (10:1), -40°+0°C, 0.5h. <sup>f</sup>AcOH/THF/H<sub>2</sub>O (3:1:1), 70°C, 3h. <sup>g</sup>10, THF/PhCH<sub>3</sub> (1:3), -78°+0°C, 2h; 0°C, 40 min. <sup>h</sup>2% HCl/MeOH, 45°C, 3h. <sup>i</sup>LiOH, MeOH/H<sub>2</sub>O (2:1), 24°C, 5h.

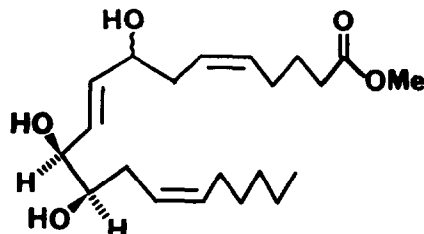
Methyl pyranoside **3**, prepared<sup>9</sup> as an anomeric mixture from commercial 2-deoxy-D-galactose (88%), was converted to **4**<sup>10</sup> (72%) by primary selective tritylation, protection of the secondary alcohols with 3,4-dimethoxybenzyl chloride, and zinc bromide mediated<sup>11</sup> detritylation (Scheme I). Swern oxidation<sup>12</sup> of **4** and condensation of the

resultant aldehyde with 1.2 equiv of 7-carbomethoxyhepta-3(Z)-en-1-ylidetriphenylphosphorane<sup>13</sup> (9) under *cis*-olefination conditions furnished ester 5 (93%) after chromatographic purification (SiO<sub>2</sub>: EtOAc/hexanes 3:1, R<sub>f</sub> ~ 0.53). The lactol obtained from 5 by mild acid hydrolysis was added dropwise in a minimum volume of toluene to a 36mM solution of hexylidetriphenylphosphorane (10) [2.7 equiv; generated at -78°C, THF, 0.5h, LiN(SiMe<sub>3</sub>)<sub>2</sub>] in toluene/THF (3:1) at -78°C. The mixture was gradually warmed to 0°C over 2h where it was maintained for an additional 40 min. Quenching with ice-cold 50% aqueous NH<sub>4</sub>OAc, extractive isolation, and flash chromatography afforded 6 (55% overall from 5) accompanied by a small amount of its 14(E)-isomer. TLC: SiO<sub>2</sub>, EtOAc/hexanes (1:1), two elutions, R<sub>f</sub> ~ 0.55 and 0.61 for 6 and 14(E)-6, respectively. Removal of the benzyl protecting groups with 2% methanolic HCl yielded methyl 10(R),11(R),12(R)-trihydroxyeicosa-5(Z),8(Z),14(Z)-trienoate<sup>14</sup> (7), [α]<sub>D</sub><sup>22</sup>-16.4° (c 3.5, acetone), which was saponified to free acid 8 (74% overall from 6); TLC: SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub> (1:9), R<sub>f</sub> ~ 0.35.



11: R = Me

12: R = H



13

Mitsunobu inversion<sup>15</sup> of 6 utilizing triphenylphosphine/diethyl azodicarboxylate/benzoic acid (2 equiv each) in THF at 0°C followed by benzoate solvolysis (NaOMe/MeOH, 22°C, 10h) and debenylation as described above generated methyl 10(S),11(R),12(R)-trihydroxyeicosa-5(Z),8(Z),14(Z)-trienoate (11), [α]<sub>D</sub><sup>22</sup>+29.2° (c 2.0, acetone). Saponification gave free acid 12 in 30% overall yield from 6; TLC: SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub> (1:9), R<sub>f</sub> ~ 0.38. Unexpectedly, triol 13, the result of allylic transposition during the Mitsunobu reaction, was also isolated and found to be an approximate 1:5 diastereomeric mixture at C(8); TLC: SiO<sub>2</sub>, MeOH/CH<sub>2</sub>Cl<sub>2</sub> (5:95), R<sub>f</sub> ~ 0.15 and 0.17.

Details of current investigations into the production and physiological relevance of trioxilins will be reported elsewhere.

**Acknowledgment:** Supported financially by grants from the USPHS NIH (GM 31278), the Robert A. Welch Foundation (I-782), and NATO (RG 85/0026). Funds for the purchase of a mass spectrometer were provided by NIH GM16488.

#### References and Notes

1. I.C. Walker, R.L. Jones, and N.H. Wilson, Prostaglandins **18**: 173-178 (1979).
2. C.R. Pace-Asciak, J. Biol. Chem. **259**: 8332-8337 (1984).
3. E.J. Corey, J. Kang, B.C. Laguzza, and R.L. Jones, Tetrahedron Letters **24**: 4913-4916 (1983).
4. C.R. Pace-Asciak, E. Granstrom, and B. Samuelsson, J. Biol. Chem. **258**: 6835-6840 (1983).
5. B.D. Hammock, M. Ratcliff, and D.A. Schooley, Life Sciences **27**: 1635-1641 (1980) and cited references.
6. C.R. Pace-Asciak, J.M. Martin, and E.J. Corey, Prog. Lipid Res. **25**: 625-628 (1986).
7. D. Piomelli, S. J. Feinmark, and J. H. Schwartz, Society for Neuroscience 17th Annual Meeting, New Orleans, Louisiana, Nov. 16-21, 1987: Abstract No. 169.11, p. 598.
8. T. Kato, Y. Yamaguchi, S.-I. Ohnuma, T. Uyehara, T. Nami, M. Kodama, and Y. Shiobara, Chemistry Letters:577-580 (1986); W.S. Powell and C.D. Funk, Prog. Lipid Res. **26**: 183-210 (1987); D.L. Holland, J. East, K.H. Gibson, E. Clayton, and A. Oldfield, Prostaglandins **29**: 1021-1029 (1985).
9. W.G. Overend, F. Shafizadeh, and M. Stacey, J. Chem. Soc.:671-677 (1950).
10. Except as noted, satisfactory spectral data were obtained for all new compounds using chromatographically homogeneous samples.
11. V. Kohli, H. Blocker, and H. Koster, Tetrahedron Letters **21**: 2683-2686 (1980).
12. A.J. Mancuso and D. Swern, Synthesis: 165-185 (1981).
13. R. Grée, H. Tourbah, and R. Carrié, Tetrahedron Letters **27**: 4983-4986 (1986).
14. Physical data for **7**:  $^1\text{H NMR}$ ( $\text{CDCl}_3$ , 300 MHz)  $\delta$  0.88 (t,  $J=6\text{Hz}$ , 3H), 1.17-1.40 (m,6H), 1.68 (dt,  $J=6$ , 12Hz, 2H), 1.95-2.15 (m,4H), 2.22-2.42 (m,4H), 2.73-3.01 (m,2H), 3.45 (dd,  $J=6$ , 6Hz, 1H), 3.65 (s,3H), 3.73 (dt,  $J=4$ , 6Hz, 1H), 4.69 (dd,  $J=4$ , 10Hz, 1H), 5.32-5.48 (m,3H), 5.52-5.74 (m,3H); MS (EI, 70ev) of TMS ether  $m/e$ : 103, 129 (base), 147, 159, 186, 213, 225, 269, 282, 315, 342; RP-HPLC:  $\mu\text{Bondapak C}_{18}$  (3.9 x 300 mm),  $\text{CH}_3\text{CN}/\text{H}_2\text{O}$  (35:65), 1 ml/min flow rate,  $R_t \sim 13.5$  min, monitored at 210 nm.  $^{11}\text{H NMR}$  ( $\text{CDCl}_3$ , 300 MHz)  $\delta$  0.88 (t,  $J=6\text{Hz}$ , 3H), 1.17-1.42 (m, 6H), 1.68 (dt,  $J=6$ , 12Hz, 2H), 1.93-2.58 (complex m, 11H), 2.74-3.03 (m, 2H), 3.42-3.55 (m, 1H), 3.56-3.72 (m with methyl ester singlet at 3.65, 4H), 4.65 (dd,  $J=6$ , 10Hz, 1H), 5.32-5.75 (m, 6H); MS (PICI,  $\text{CH}_4$ ) of TMS ether  $m/e$ : 213 (base), 269, 287, 315, 343, 355, 405, 429, 479, 511, 555, 569, 585.
15. O. Mitsunobu, Synthesis: 1-28 (1981).

(Received in USA 11 March 1988)